

CHROM 12,290

Note

Gas chromatographic separation of the methyl ether methylglycopyranoside series hexose, 6-deoxyhexose and pentose acetates

Yu. N. ELKIN

Pacific Institute of Bio-Organic Chemistry, Far East Science Centre, Academy of Sciences of the U.S.S.R., Vladivostok-22 (U.S.S.R.)

(Received July 30th, 1979)

The derivatives used most widely for determining the positions of glycosidic linkages in sugar residues in polysaccharides are partially methylated alditol acetates¹. They offer information on glycosidic linkages by means of their mass spectra, while the kind of monosaccharide epimer can be determined by its gas-liquid chromatographic (GLC) retention index (R_t). Derivatives of this type have some limitations. Some of them, owing to the overlapping of GLC peaks, have been overcome by using ethyl ethers instead of methyl ethers² or by additional separation using thin-layer chromatography³. However, the alditol acetates can be applied only with the aldose- and ketose-containing polysaccharides. A general method for investigating substances containing hexosamine, N-acetylneuraminic acid (NANA) and uronic acid, involves methanolysis. This reaction has the advantage over hydrolysis of preventing the extensive destruction of neutral sugars^{4,5}. The double number of methyl ethers in the methanolysis products is likely to complicate their chromatographic separation owing to the formation of α - and β -anomers. On the other hand, each sugar is characterized by four GLC values, *viz.*, two R_t values for the anomers, and their partial peak areas (R_a).

To increase the volatility, partially methylated sugars are converted into trimethylsilyl ethers or acetylated derivatives. The latter seem preferable for two reasons. Firstly, the mass spectra of this type of derivative of aldoses⁶, hexosamine⁷⁻¹⁰, NANA^{11,12} and uronic acids¹³ have been extensively studied. Secondly, the acetate derivatives are capable of being stored. Also, one can readily obtain samples of methyl ether methylglycosides by partial methylation of the methylglycoside required^{5,14,15}. However, the available R_t data of some methyl ethers, methyl α - and β ,L-Ara¹⁶, α - and β ,D-Glc¹⁷, β ,D-Xyl¹⁵ and α ,D-Gal⁵ acetate are not sufficient for the wide use of these derivatives in analysing the methylation of aldose-containing polysaccharides. This paper presents retention indices for the required set of methyl ether acetates of the following methylglycosides: α - and β ,D-Glc, α - and β ,D-Gal, α ,D-Man, α - and β ,D-Qui, α - and β ,L-Fuc, α ,D-Rha, α - and β ,D-Xyl, α - and β ,L-Ara and α ,D-Lyx.

EXPERIMENTAL

A method of preparing the required set of methyl ether methylglycosides has already been described¹⁴. The GLC separation was performed on Pye Unicam 104 GLC and LKB 9000s GLC-MS instruments. In determining R_t values we used columns, liquid phases and operating conditions as follows: 200 × 0.4 cm I.D. glass column containing 3% QF-1 (A) and 3% NPGS (B) on Chromosorb W (100-120 mesh). Columns A and B were temperature programmed at 5°/min from 110° to 230°. Argon was used as the carrier gas at a flow-rate of 60 ml/min. The GLC-MS system was operated with 1% liquid phase column and with helium as the carrier gas at a flow-rate of 25 ml/min. The R_t scale was O-methyl-2,3,4-O-trimethyl- β ,D-Xyl; 1000-aldonitrile of D-Gal acetate.

Three types of measurement were made. To a first approximation an R_t value was determined by injecting into the column a set of methyl ethers of each methylglycoside, dissolved in chloroform secondly, to determine the relative R_t , two sets of methyl ethers of the same type of methylglycoside (e.g., hexosides) were injected into the column simultaneously in different proportions. Thirdly, if the peaks of the methyl ethers coincided, they were identified by studying the mass spectra taken over the GLC peak contour.

RESULTS AND DISCUSSION

The R_t data are given in Tables I-VIII according to the monosaccharide ring methylation pattern. In each table, the list of sugars is divided into epimer groups as the weight of the C-5 substituent increases. Within each group, the epimers listed follow one another as the R_t value of the α -anomer increases, followed by the β -

TABLE I
RETENTION INDICES OF Me 2,3,4-OMe GLYCOPYRANOSIDES

<i>C-5 substituent</i>	<i>Anomer</i>	<i>Sugar</i>	<i>QF-1</i>	<i>NPGS</i>
H	α	Xyl	031	039
	β	Xyl	000	000
	α	Lyx	055	057
	α	Ara	116	128
	β	Ara	112	128
CH ₃	α	Qui		000
	β	Qui		030
	α	Rha	016	008
	α	Fuc	085	078
	β	Fuc	074	063
CH ₂ OMe	α	Glc	183	190
	β	Glc	125	123
	α	Man	183	190
	α	Gal	200	230
	β	Gal	200	225
CH ₂ OAc	α	Glc	407	417
	β	Glc	352	360
	α	Man	407	436
	α	Gal	442	494

TABLE II

RETENTION INDICES OF Me 2,3-OMe₂ GLYCOPYRANOSIDE ACETATES

<i>C-5 substituent</i>	<i>Anomer</i>	<i>Sugar</i>	<i>QF-1</i>	<i>NPGS</i>
H	α	Ara	202	254
	β	Ara	220	254
	α	Xyl	270	265
	β	Xyl	220	203
	α	Lyx	282	286
CH ₃	α	Fuc	200	193
	β	Fuc	172	181
	α	Qui	257	209
	β	Qui	222	170
CH ₂ OMe	α	Rha	287	222
	α	Gal	352	399
	β	Gal	324	371
	α	Glc	436	436
	β	Glc	385	371
CH ₂ OAc	α	Man	448	449
	α	Gal	539	589
	β	Gal	502	560
	α	Glc	620	640
	β	Glc	585	584
	α	Man	639	640

TABLE III

RETENTION INDICES OF Me 2,4-OMe₂ GLYCOPYRANOSIDE ACETATES

<i>C-5 substituent</i>	<i>Anomer</i>	<i>Sugar</i>	<i>QF-1</i>	<i>NPGS</i>
H	α	Ara	230	242
	β	Ara	336	323
	α	Lyx	294	286
	α	Xyl	398	350
	β	Xyl	305	286
CH ₃	α	Rha	260	193
	α	Fuc	340	286
	β	Fuc	289	237
	α	Qui	365	286
CH ₂ OMe	β	Qui	280	214
	α	Man	436	416
	α	Gal	495	494
	β	Gal	448	449
	α	Glc	557	518
CH ₂ OAc	β	Glc	466	436
	α	Man	654	640
	α	Gal	722	697
	β	Gal	675	668
	α	Glc	769	743
	β	Glc	681	655

anomer. This method of presentation of R_i is convenient for the identification of epimers when the methylation pattern, and sometimes that of anomers, is determined from the mass spectrum. The β -anomer of the mannose series and thirteen compounds

TABLE IV
RETENTION INDICES OF Me 3,4-OMe₂ GLYCOPYRANOSIDE ACETATES

C-5 substituent	Anomer	Sugar	QF-1	NPGS
H	α	Lyx	168	160
	α	Xyl	199	167
	α	Ara	353	355
CH ₃	α	Rha	125	115
	α	Qui	182	160
	α	Fuc	287	237
CH ₂ OMe	α	Man	284	262
	α	Glc	375	314
	α	Gal	426	424
CH ₂ OAc	α	Man	524	505
	α	Glc	614	584
	α	Gal	675	647

TABLE V
RETENTION INDICES OF Me 2-OMe GLYCOPYRANOSIDE ACETATES

C-5 substituent	Anomer	Sugar	QF-1	NPGS
H	α	Ara	412	400
	β	Ara	500	460
	α	Lyx	508	445
	α	Xyl	518	450
	β	Xyl	430	385
CH ₃	α	Rha	470	364
	α	Qui	479	385
	β	Qui	395	311
	α	Fuc	486	404
	β	Fuc	491	358
CH ₂ OMe	α	Man	624	589
	α	Glc	644	622
	β	Glc	565	535
	α	Gal	648	622
	β	Gal	565	571
CH ₂ OAc	α	Man	790	766
	α	Gal	803	783
	β	Gal	728	734
	α	Glc	814	797
	β	Glc	732	734

in the set of methyl ethers¹⁴ were not investigated as they were not formed under the conditions of methanolysis used. The last conclusion was confirmed when determining the positions of glycoside linkages of sugars in a lipopolysaccharide from *Yersinia pseudotuberculosis*^{18,19}.

Numerical values of the retention indices were measured as a first approximation to within ± 10 for methyl ethers of the three selected methylglycosides, *i.e.*, β ,D-Xyl, α ,L-Fuc and α ,D-Man. The relative R_t values of epimers in each group were then determined by GLC, making measurements to within ± 5 on pairs of single type sugar ether sets. For example (Table III), the R_t value equal to 340 for methyl

TABLE VI

RETENTION INDICES OF Me 3-OMe GLYCOPYRANOSIDE ACETATES

<i>C-5 substituent</i>	<i>Anomer</i>	<i>Sugar</i>	<i>QF-1</i>	<i>NPGS</i>
H	α	Lyx	398	378
	α	Xyl	454	400
	β	Xyl	512	458
	α	Ara	464	460
	β	Ara	387	400
CH ₃	α	Rha	353	284
	α	Fuc	365	331
	β	Fuc	405	338
	α	Qui	450	351
	β	Qui	542	338
CH ₂ OMe	α	Man	517	505
	α	Gal	517	528
	β	Gal	620	642
	α	Glc	614	589
	β	Glc	684	645
CH ₂ OAc	α	Man	710	697
	α	Gal	705	715
	β	Gal	792	797
	α	Glc	803	772
	β	Glc	866	821

TABLE VII

RETENTION INDICES OF Me 4-OMe GLYCOPYRANOSIDE ACETATE :

<i>C-5 substituent</i>	<i>Anomer</i>	<i>Sugar</i>	<i>QF-1</i>	<i>NPGS</i>
H	α	Lyx	476	400
	α	Xyl	486	405
	β	Xyl	466	429
	α	Ara	486	439
CH ₃	α	Rha	445	306
	α	Qui	470	335
	β	Qui	460	358
	α	Fuc	502	385
CH ₂ OMe	α	Man	601	528
	β	Glc	616	584
	β	Gal	686	642
CH ₂ OAc	α	Man	829	743
	α	Glc	859	792
	β	Glc	836	797
	α	Gal	866	788
	β	Gal	894	853

2,4-O-dimethyl-3-O-acetyl- α ,L-fucopyranoside lies within the range 330–350, but methyl 2,4-O-dimethyl-3-O-acetyl- β ,D-quinovopyranoside ($R_t = 355$) elutes after the former from column A, the R_t value being higher by 15 ± 5 .

The double number of methyl ethers of each monosaccharide reduced the possibility of complete separation. For this reason, epimer groups have some unseparated pairs, as shown in Table IX. However, the problem of whether one or two isomers

TABLE VIII
RETENTION INDICES OF Me GLYCOPYRANOSIDE ACETATES

<i>C-5 substituent</i>	<i>Anomer</i>	<i>Sugar</i>	<i>QF-1</i>	<i>NPGS</i>
H	α	Xyl	622	506
	β	Xyl	630	550
	α	Lyx	617	523
	α	Ara	617	565
	β	Ara	602	518
CH ₃	α	Rha	562	436
	α	Qui	570	443
	β	Qui	590	480
	α	Fuc	582	461
CH ₂ OMe	β	Fuc	605	525
	α	Man	710	640
	α	Gal	722	665
	β	Gal	745	733
CH ₂ OAc	α	Glc	740	670
	β	Glc	732	705
	α	Man	882	818
	α	Gal	888	828
—	β	Gal	894	905
	α	Glc	899	853
	β	Glc	902	898

TABLE IX
POORLY SEPARATED PAIRS OF ISOMERIC METHYL ETHER METHYL GLYCOPYRANOSIDE ACETATES

<i>Positions of OMe groups</i>	<i>Anomer</i>	<i>Pairs of isomeric sugar</i>	<i>Difference in R_t values on different columns</i>	
			<i>QF-1</i>	<i>NPGS</i>
2, 3, 4, 6	α	Glc	0	0
	α	Man		
2, 3, 4, 6	α	Gal	0	5
	β	Gal		
2, 3, 4	α	Ara	4	0
	β	Ara		
2, 6	α	Glc	4	0
	α	Gal		
2	β	Gal	4	0
	β	Glc		
4	β	Gal	7	4
	β	Glc		
—	α	Rha	8	7
	α	Qui		
—	β	Gal	8	7
	β	Glc		

constitute a particular GLC peak can be solved by finding out the second separated anomers for the pair under suspicion. For example, GLC-MS analysis of the acetylated methanolysis products of any polysaccharide reveals three peaks of the Me 2,6-OMe₂-3,6-OAc₂-hexopyranosides which lie within the R_t ranges 525-555, 560-580 and 610-630 on the NPGS column. Differences between them are 12 ± 5

and 35 ± 5 . Thus, α anomers of glucose and galactose derivatives compose the third peak, the first and second peaks are β anomers of glucose and galactose derivatives, respectively, as shown in Table V.

In conclusion, Tables I–VIII, giving the retention indices of methyl ether methylglycoside acetates, provide a reliable means of determining the epimers of the sugar series studied in the GLC–MS analysis. Out of 285 pairs of epimer sets, there are only eight unseparated pairs.

REFERENCES

- 1 H. Björndal, C. G. Hellerqvist, B. Lindberg and S. Svenson, *Angew. Chem.*, (1970) 634.
- 2 D. P. Sweet, P. Albersheim and R. H. Shapiro, *Carbohydr. Res.*, 40 (1975) 199.
- 3 Y.-M. Choy, G. G. S. Dutton, K. B. Gibney, S. Kabir and J. N. C. Whyte, *J. Chromatogr.*, 72 (1972) 13.
- 4 G. R. Jamieson and E. H. Reid, *J. Chromatogr.*, 101 (1974) 185.
- 5 J. Montreuil, *Pure Appl. Chem.*, 42 (1975) 435
- 6 Yu. N. Elkin, B. V. Rozinov, A. I. Kalinovsky, A. F. Pavlenko, N. I. Shulga and A. K. Dzizenko, *Khim. Prirod. Soed.*, (1973) 601.
- 7 K. Heyns, G. Kiessling and D. Muller, *Carbohydr. Res.*, 4 (1967) 452.
- 8 A. I. Kalinovsky and A. K. Dzizenko, *Khim. Prirod. Soed.*, (1974) 780.
- 9 S. K. Kundu and R. W. Ledeen, *Carbohydr. Res.*, 39 (1975) 179.
- 10 A. Stoffyn, P. Stoffyn and J. C. Orr, *Carbohydr. Res.*, 23 (1972) 251.
- 11 H. Rauvala and J. Kärkkäinen, *Carbohydr. Res.*, 56 (1977) 1.
- 12 H. von Holbeek, J. P. Kamerling and J. F. G. Vliegthart, *Carbohydr. Res.*, 60 (1978) 51.
- 13 V. Kovačik, V. Michalov and P. Kovač, *Carbohydr. Res.*, 54 (1977) 23.
- 14 Yu. N. Elkin, N. I. Shulga, T. I. Vakorina and A. K. Dzizenko, *Anal. Biochem.*, 68 (1975) 9.
- 15 E. V. Evtushenko and Yu. S. Ovodov, *J. Chromatogr.*, 97 (1974) 99.
- 16 S. C. Williams and J. K. N. Jones, *Can. J. Chem.*, 45 (1967) 275.
- 17 H. G. Jones and J. K. N. Jones, *Can. J. Chem.*, 47 (1969) 3269.
- 18 S. V. Tomshich, R. P. Gorshkova, Yu. N. Elkin and Yu. S. Ovodov, *Khim. Prirod. Soed.*, (1975) 563.
- 19 S. V. Tomshich, R. P. Gorshkova, Yu. N. Elkin and Yu. S. Ovodov, *Eur. J. Biochem.*, 65 (1976) 193.